
Academic Year: 2018-2019

Course: DIAGNOSTIC MEDICAL BIOTECHNOLOGY

TYPE OF EDUCATIONAL ACTIVITY :

Teacher: Prof. Angela OSTUNI

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Web site

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mobile:

Language: ITALIAN

ECTS: 6**(5 of lessons and 1 of tutorials/practice)****n. hours: 52****(40 of lessons and 12 of tutorials/practice)****Campus: Potenza****Dept: Sciences****Program:BIOTECNOLOGIE (L2)****Semester: I****(from 01/10/2018 to 20/12/2018-20/01/2019)**

EDUCATIONAL GOALS AND EXPECTED LEARNING OUTCOMES

- Know and be able to use technological systems to be applied in the context of human health
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PRE-requirements

- Students must have acquired the knowledge of General Pathology and Molecular Biology
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Syllabus

- ✓ Production of recombinant proteins in heterologous systems used in the medical field.
 - ✓ Transgenic animals and plants for drugs production.
 - ✓ Gene therapy
 - ✓ Antigens, immunogens, epitopes. Major histocompatibility complex. Inactivated, attenuated, vaccines subunits. Adjuvants. Strategies for developing vaccines without the use of adjuvants. DNA vaccines. Passive immunoprophylaxis.
 - ✓ Design of vaccines for HIV and cancer
 - ✓ Preparation of polyclonal antibodies and their application
 - ✓ Preparation of monoclonal antibodies: hybridoma method; humanization of antibodies; phage display.
 - ✓ Monoclonal antibodies in therapy: preparation of immune conjugates (toxins, radionuclides, drugs, enzymes) immune liposomes, immune polimers.
 - ✓ Antibodies in diagnostic field: ELISA, western blotting, Immunocytochemistry, immunohistochemistry, immunofluoresce, flow cytometry
 - ✓ Cell cultures: technical set-up and maintenance
 - ✓ Stem cells: sources, isolation and their use in clinic
 - ✓ Cell Therapy
 - ✓ Molecular therapy: mechanisms of signal transduction, strategies to block cell surface receptors and ligand, protein kinase inhibitors; inhibitors of the enzyme farnesyltransferase, inhibitors of angiogenesis. Metallo protease inhibitors.
 - ✓ Oligonucleotides as molecular probes: the fluorescence in situ hybridization (FISH).
 - ✓ Oligonucleotides as therapeutic agents: antisense oligonucleotides, RNA interference.
 - ✓ Assisted reproductive technologies.
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TEACHING METHODS

- The course includes 40 hours of classroom teaching and 12 hours of guided exercises in the laboratory e/or in the classroom.
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EVALUATION METHODS

Oral exam on all course topics and critical discussion of the experience in the laboratory. The student must submit a written report on the lab experience one week before the examination date.

TEXTBOOKS AND EDUCATIONAL MATERIAL

- Metodologie di base per le scienze biomolecolari. Rob Reed, David Holmes, Jonathan Weyers, Allan Jones. Zanichelli
 - Introduzione alle colture cellulari. Mariottini et al. Morgan Edizioni tecniche
 - Le cellule staminali e la terapia genica. Antonino Sapuppo. Giunti
 - Biologia cellulare e genetica. Parte I (cap.16, 18,21). A. Fantoni et al. Ed. Piccin
 - Fondamenti di immunologia. A.K. Abbas, A.H. Lichtman. Ed Piccin
 - Introduzione alla medicina molecolare. D.W. Ross. Springer ed.
 - Teacher's slides
 - scientific articles on specific topics
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INTERACTION WITH STUDENTS

At the beginning of the course, after describing the objectives, the detailed program and methods of verification, the teacher will indicate the reference texts and the availability of teaching materials. Teacher will collect the list of students, together with name, serial number and email. Teacher will be available for contact with the students by appointment fixed through e-mail .

EXAMINATION SESSIONS

February 26, March 26, June 11, July 16, September 24, October 29, December 10

SEMINARS BY EXTERNAL EXPERTS SI NO

FURTHER INFORMATION
